

Evaluation of Micro-Electrocorticographic Electrodes for Electrostimulation

Seth J. Wilks, Andrew S. Koivuniemi, Sanitta Thongpang, Justin C. Williams, *Member, IEEE*, and Kevin J. Otto, *Member, IEEE*

Abstract—Chronic neural recording and stimulation on the surface of the cortex with macroelectrodes has been shown to be promising for treating a wide range of neurological deficits. To enhance the specificity of these devices, dense arrangements of small area electrodes have been microfabricated for precise recording and control of neural populations. In this study micro-electrocorticographic (μ ECoG) electrodes were evaluated for electrostimulation. Surface modification with electrodeposited iridium oxide (EIrOx) resulted in lower impedance, higher charge carrying capacity, and lower, more linear voltage excursions during current controlled stimulation.

I. INTRODUCTION

NEUROPROSTHETIC brain-computer interface (BCI) systems have received interest in the neuroengineering field for treatment of some neurological injuries and disorders. Electrococtogram (ECoG) recordings taken from electrodes on the surface of the cortex have been successfully implemented for BCI control [1-3] and more recently μ ECoG devices have been fabricated with a dense arrangement of small-area microelectrodes on the order of 100s of microns for increased spatial resolution [4]. While these devices are commonly studied for output-type neural prosthetic systems, input-type systems may also be applicable to treat neurological problems including sensory deficits, Parkinson's disease, epilepsy, and depression.

Electrical stimulation on the surface of the primary sensory regions of the cortex has long been shown to evoke sensation [5, 6]. A group of patients who received the first cortical prostheses stimulating the surface of the primary visual cortex displayed sensory thresholds typically between 1-5 mA, which was not dependent on electrode size [7]. With the goal to decrease stimulation thresholds and increase spatial resolution, smaller, penetrating electrodes also have

been used successfully in delivering behaviorally relevant information [8-11]. The major disadvantage of penetrating the cortical surface is the tissue response associated with insertion and the chronic presence of the device [12, 13].

The main problems with using μ ECoG devices for electrostimulation result from delivering current on the order of a few microamps through high impedance sites with diameters on the order of 100s of microns. Delivering such charge densities requires relatively high voltages which can result in toxic redox reactions detrimental to both the electrode and tissue. Improvements in charge transfer have been shown through the application of different surface modifying materials such as iridium oxide [14], poly(3,4-ethylene dioxythiophene) or PEDOT [15, 16], and carbon nanotubes (CNTs) [17].

This report evaluates flexible polyimide μ ECoG devices for electrostimulation. Bare platinum electrodes and platinum electrodes modified with electrodeposited iridium oxide were characterized through electrochemical impedance spectroscopy (EIS), cyclic voltammetry (CV), and voltage excursion measurements in saline

II. MATERIALS AND METHODS

A. μ ECoG Electrode Arrays

Fabrication of μ ECoG electrode arrays has been described in detail elsewhere [4]. In brief, electrode sites made with

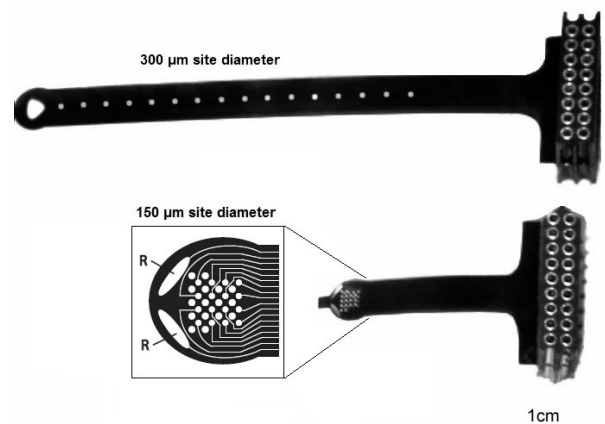


Fig. 1. μ ECoG electrode arrays. The 1x16 array on top contains large site sizes (300 μ m diameter). The 4x4 array on bottom is better equipped for *in vivo* applications with small site sizes (150 μ m diameter), two large area reference (R) sites, and holes to promote consistent perfusion.

Manuscript received April 23, 2009. This work was supported in part by the Purdue Research Foundation.

S. J. Wilks is with the Weldon School of Biomedical Engineering, Purdue University, West Lafayette, IN 47907 USA (e-mail: swilks@purdue.edu).

A. S. Koivuniemi is with the Weldon School of Biomedical Engineering and the Indiana University School of Medicine, Indianapolis, IN 46202 USA (e-mail: akoivuni@iupui.edu).

S. Thongpang and J. C. Williams are with the Biomedical Engineering Department, University of Wisconsin, Madison, WI 53706 USA (e-mail: jwilliams@engr.wisc.edu).

K. J. Otto is with the Weldon School of Biomedical Engineering and the Department of Biological Sciences, Purdue University, West Lafayette, IN 47907 USA (phone: 765-496-1012; fax: 765-496-1912; e-mail: kotto@purdue.edu).

200 nm of gold and 10 nm of platinum on top were plated onto a flexible polyimide platform. μ ECoG electrodes used in this study include a 1x16 array of 300 μ m diameter sites spaced 1.5 mm apart and a 4x4 array of 150 μ m diameter sites spaced 400 μ m apart as seen in fig. 1. The 4x4 array equipped with holes to promote consistent perfusion and two large area reference electrodes is well suited for implantation over the rat cortex.

B. Electrochemical Characterization

Electrochemical measurements were made using an Autolab Potentiostat/Galvonostat PG-STAT12 (Eco Chemie, Utrecht, Netherlands) with a built-in frequency response analyzer (Brinkmann, Westbury, NY). A three-electrode cell configuration in 1X phosphate buffered saline (PBS) was used. The microelectrode site, a large-area platinum wire, and an Accumet, gel-filled, KCl saturated calomel electrode (Thermo Fischer Scientific, Fair Lawn, NJ) functioned as the working electrode (WE), counter electrode (CE), and reference electrode.

EIS measurements were collected by configuring the Autolab to sequentially inject overpotentials of 5 mV_{RMS} sine waves at 36 frequencies logarithmically spaced from 1 Hz to 1 MHz. CV measurements were collected by configuring the Autolab to sweep the voltage of the WE between +0.8 V and -0.6 V at 50 mV/s and monitor the resulting current flow between the WE and CE. The cathodal charge storage capacity (Q_{cap}) was calculated by integrating the cathodal current density enclosed by the CV and dividing by the sweep rate.

Voltage excursion measurements were collected during the delivery of constant current pulses in 1X PBS. Cathodal-first, charge-balanced, biphasic current pulses were applied between the microelectrode site and a large-area platinum wire. 500 μ s phase durations and 100 μ A – 1 mA current amplitudes were applied at a pulsing frequency of 50 Hz. The voltage excursions were measured with respect to the platinum wire.

C. Electrodeposited iridium oxide (EIrOx)

Iridium oxide was electrodeposited in a solution of 4 mM IrCl₄, 40 mM oxalic acid, and 340 mM K₂CO₃ using a previously documented protocol [14]. In brief, the potential was first cycled 50 times between 0.0 V and 0.55 V at a sweep rate of 50 mV/s followed by cathodic and anodic biasing at the same potential limits for 1600 seconds at a pulsing frequency of 1 Hz.

III. RESULTS AND DISCUSSION

A. Electrochemical Characterization

EIS data were collected in PBS between 1Hz and 1 MHz for the two μ ECoG electrode sizes. Fig. 2a shows the impedance magnitude values were lower for the larger electrode sizes across all frequencies. The 1 kHz impedance magnitudes are of particular interest since the fundamental

frequency of an action potential is approximately 1 kHz. The 150 μ m diameter electrodes displayed a 1 kHz impedance magnitude of approximately 100 k Ω which was about one order of magnitude greater than that of sites double in diameter.

CV measurements were collected to evaluate the presence of electrochemical reactions, the double-layer capacitance, and the charge available at the interface (Q_{cap}). A slow sweep rate (50 mV/s) was used to assess the near equilibrium voltage/current relationships. As seen in fig. 2b, the bare platinum interface displays relatively smooth CVs and low Q_{cap} values less than 1 mC/cm². For stimulation applications only a small percentage of the equilibrium Q_{cap} can be accessed during fast current pulsing. For this reason, bare platinum is not suitable for safely delivering current on the order of milliamps, and surface modification is often necessary to increase the electrochemical surface area and improve the charge transfer properties of the electrode-electrolyte interface.

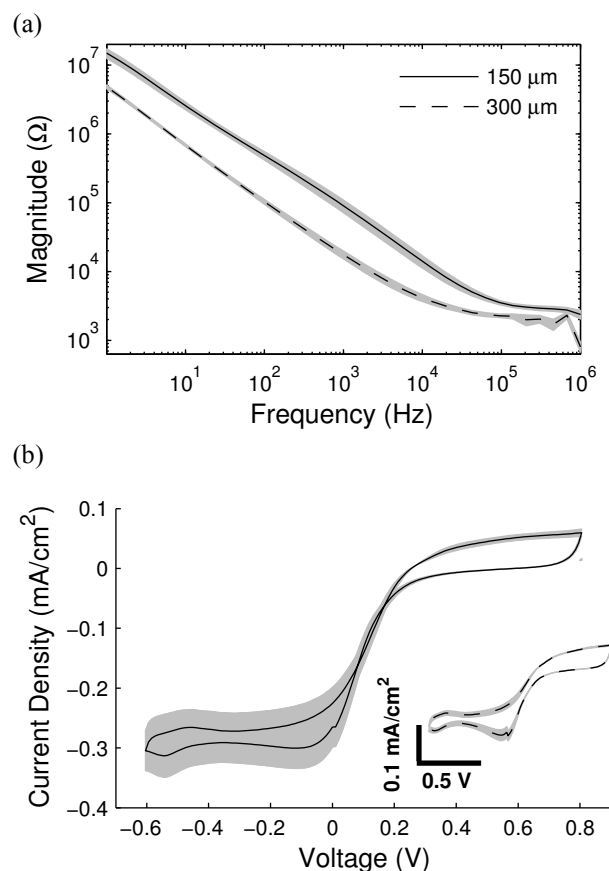


Fig. 2. Electrochemical characterization; (a) mean impedance magnitude for 36 frequencies logarithmically spaced between 1 Hz and 1 MHz, (b) mean CVs taken between -0.6 V and +0.8 V with a sweep rate of 50 mV/s for 150 μ m diameter sites (insert: 300 μ m diameter sites). The grey areas represent standard error.

B. Electrodeposited iridium oxide (EIrOx)

Iridium oxide has commonly been used to modify the surface of microelectrodes for electrostimulation and can easily be activated on iridium electrodes or electrodeposited

on other metal surfaces [14]. EIrOx increases the electrochemical surface area of the electrode and improves charge transfer through electron exchange that occurs during reversible oxidation and reduction reactions.

As expected, application of EIrOx to the μ ECoG electrode sites significantly improved the interface for charge transfer represented by the larger CV shown in fig. 3a. The EIrOx modified sites displayed characteristic peaks associated with the reversible oxidation and reduction of iridium. EIrOx greatly increased the charge available at the interface corresponding to an increase in Q_{cap} to 34.2 mC/cm^2 for the $150 \mu\text{m}$ diameter sites and to 14.8 mC/cm^2 for the $300 \mu\text{m}$ diameter sites (fig. 3b). The same iridium oxide deposition duration was applied for both electrode sizes which explains why the smaller electrodes displayed a larger Q_{cap} . Interestingly, the 1 kHz impedance magnitude dropped to similar values for the two site sizes: $2.7 \text{ k}\Omega$ for the $300 \mu\text{m}$ electrode sites and $3.0 \text{ k}\Omega$ for the $150 \mu\text{m}$ electrode sites (fig. 3c).

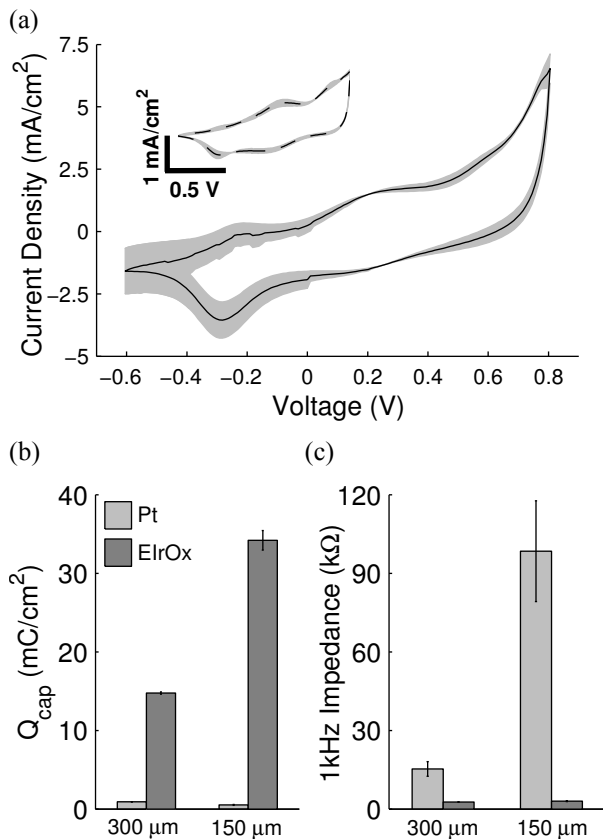


Fig. 3. EIrOx modification; (a) $150 \mu\text{m}$ diameter sites coated with EIrOx showed an increased CV curve with peaks characteristic of iridium oxide (insert: $300 \mu\text{m}$ diameter sites). This resulted in increased (b) Q_{cap} and decreased (c) 1 kHz impedance magnitude. Grey regions and error bars represent standard error.

Monitoring the amplitude and shape of the resultant voltage excursions during current controlled stimulation is often done to evaluate the safety of the interface during charge transfer. A common limitation is operating within the potentials which result in hydrolysis. Limits commonly used

for platinum and iridium oxide electrodes are $+0.8 \text{ V}$ and -0.6 V . Exceeding these limits could result in trauma due to changes in pH as hydrogen and hydroxide ions build up.

$300 \mu\text{m}$ diameter sites were pulsed at biologically relevant parameters at an amplitude of 1 mA and a phase duration of $500 \mu\text{s}$ before and after application of EIrOx. EIrOx modified sites displayed a lower amplitude voltage excursion as seen in Fig. 4a. Further, the EIrOx voltage excursion exhibited a much more ohmic representation of the applied current pulse.

Because of the voltage compliance of the stimulator device, the current amplitude was reduced to $100 \mu\text{A}$ for the $150 \mu\text{m}$ diameter sites as seen in fig. 4b. Similar to that seen with the larger electrodes, the EIrOx modified surface displayed a more linear, low amplitude voltage excursion. Prior to EIrOx modification the platinum voltage excursions exceeded the limits of hydrolysis during $100 \mu\text{A}$ pulsing. The EIrOx modified electrode operated within $\pm 0.2 \text{ V}$, well within the limits of hydrolysis. Additionally, the stimulator was able to operate within its compliance voltage during application of 1 mA current pulses.

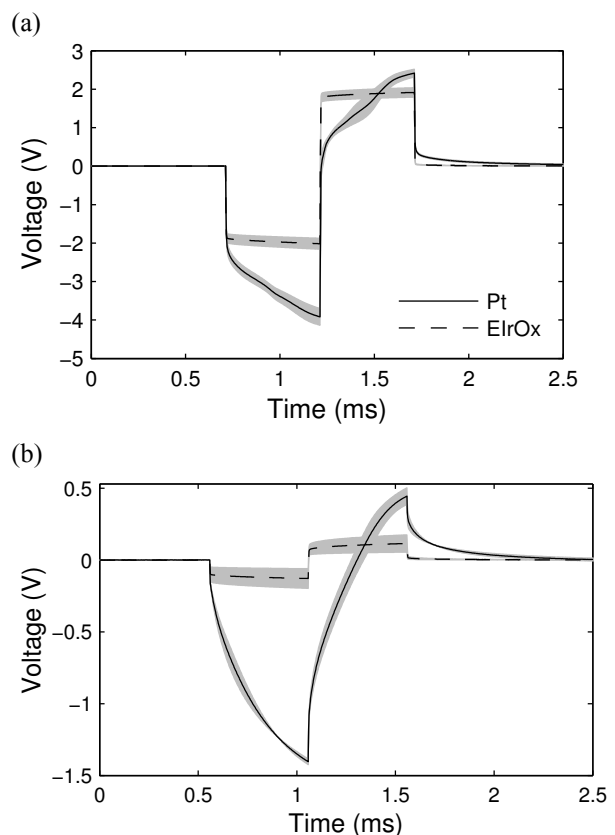


Fig. 4. Voltage excursions; (a) 1 mA current pulses were delivered through the $300 \mu\text{m}$ diameter sites and (b) $100 \mu\text{A}$ current pulses were delivered through the $150 \mu\text{m}$ diameter sites. EIrOx decreased the voltage excursion amplitude and displayed a more ohmic representation of the current pulse. Grey areas represent standard error.

μ ECoG systems present a unique neuroprosthetic platform between devices placed outside the skull and those penetrating the brain. They can provide more selective

stimulation than transcranial stimulation and reduce short- and long-term complications with electrode placement and presence because they are less invasive than penetrating arrays. The main concern with cortical surface stimulation with electrodes on the order of 100s of microns in diameter is the ability to operate at large charge densities without causing tissue trauma. While iridium oxide greatly improves charge transfer properties, its long-term stability may be a problem. Delivering current pulses at an amplitude of 1 mA and phase duration of 500 μ s with 150 μ m diameter electrodes results in a charge density of approximately 3 mC/cm². Application of such charge densities has been shown to cause iridium oxide instability resulting in damage to both the electrode and tissue [18, 19].

Further exploration into long-term electrochemical stability and efficacy in neural activation must still be done. Additionally, because of the relatively large amplitudes required for cortical surface stimulation, alternative coatings with better charge transfer properties than iridium oxide must be developed. Materials such as PEDOT and CNTs have recently emerged as potential materials for neural stimulation and have successfully been applied to electrode surfaces [15-17]. PEDOT coatings have been reported as having a Q_{cap} as much as three times greater than that seen with iridium oxide [20] and CNTs have become very popular in the miniaturization of electrical devices. Additionally, these materials are attractive for biological applications because they can be functionalized with bioactive molecules. Further work on improving the stability of these coating must be done to validate their chronic application.

IV. CONCLUSION

μ ECoG platinum electrode arrays were evaluated for electrostimulation. IrOx modified electrodes displayed a low impedance magnitude and a high Q_{cap} . During constant current stimulation, IrOx modified electrodes exhibited low amplitude and linear voltage excursions. The data presented in this report suggest surface modified μ ECoG electrodes are practical for electrostimulation applications.

ACKNOWLEDGMENT

The authors would like to thank the Purdue NeuroProstheses Research lab for helping in the preparation of this manuscript.

REFERENCES

- [1] E. C. Leuthardt, G. Schalk, J. R. Wolpaw, J. G. Ojemann, and D. W. Moran, "A brain-computer interface using electrocorticographic signals in humans," *J. Neural Eng.*, vol. 1, pp. 63-71, Jun 2004.
- [2] G. Schalk, K. J. Miller, N. R. Anderson, J. A. Wilson, M. D. Smyth, J. G. Ojemann, D. W. Moran, J. R. Wolpaw, and E. C. Leuthardt, "Two-dimensional movement control using electrocorticographic signals in humans," *J. Neural Eng.*, vol. 5, pp. 75-84, Mar 2008.
- [3] J. A. Wilson, E. A. Felton, P. C. Garell, G. Schalk, and J. C. Williams, "ECoG factors underlying multimodal control of a brain-computer interface," *IEEE Trans. Neural Syst. Rehabil. Eng.*, vol. 14, pp. 246-50, Jun 2006.

- [4] J. Kim, J. A. Wilson, and J. C. Williams, "A cortical recording platform utilizing microECoG electrode arrays," *Conf. Proc. IEEE Eng. Med. Biol. Soc.*, vol. 2007, pp. 5353-7, 2007.
- [5] G. S. Brindley and W. S. Lewin, "The sensations produced by electrical stimulation of the visual cortex," *J. Physiol.*, vol. 196, pp. 479-93, May 1968.
- [6] W. Penfield and P. Perot, "The Brain's Record of Auditory and Visual Experience. a Final Summary and Discussion," *Brain*, vol. 86, pp. 595-696, Dec 1963.
- [7] W. H. Dobbelle and M. G. Mladejovsky, "Phosphenes produced by electrical stimulation of human occipital cortex, and their application to the development of a prosthesis for the blind," *J. Physiol.*, vol. 243, pp. 553-76, Dec 1974.
- [8] M. J. Bak, J. P. Girvin, F. T. Hambrecht, C. V. Kufta, G. E. Loeb, and E. M. Schmidt, "Visual sensations produced by intracortical microstimulation of the human occipital cortex," *Med. Biol. Eng. Comput.*, vol. 28, pp. 257-9, May 1990.
- [9] K. J. Otto, P. J. Rousche, and D. R. Kipke, "Microstimulation in auditory cortex provides a substrate for detailed behaviors," *Hear. Res.*, vol. 210, pp. 112-7, Dec 2005.
- [10] K. J. Otto, P. J. Rousche, and D. R. Kipke, "Cortical microstimulation in auditory cortex of rat elicits best-frequency dependent behaviors," *J. Neural Eng.*, vol. 2, pp. 42-51, Jun 2005.
- [11] P. J. Rousche, K. J. Otto, M. P. Reilly, and D. R. Kipke, "Single electrode micro-stimulation of rat auditory cortex: an evaluation of behavioral performance," *Hear. Res.*, vol. 179, pp. 62-71, May 2003.
- [12] K. J. Otto, M. D. Johnson, and D. R. Kipke, "Voltage pulses change neural interface properties and improve unit recordings with chronically implanted microelectrodes," *IEEE Trans. Biomed. Eng.*, vol. 53, pp. 333-40, Feb 2006.
- [13] J. C. Williams, J. A. Hippensteel, J. Dilgen, W. Shain, and D. R. Kipke, "Complex impedance spectroscopy for monitoring tissue responses to inserted neural implants," *J. Neural Eng.*, vol. 4, pp. 410-23, Dec 2007.
- [14] R. D. Meyer, S. F. Cogan, T. H. Nguyen, and R. D. Rauh, "Electrodeposited iridium oxide for neural stimulation and recording electrodes," *IEEE Trans. Neural Syst. Rehabil. Eng.*, vol. 9, pp. 2-11, Mar 2001.
- [15] X. T. Cui and D. D. Zhou, "Poly (3,4-Ethylenedioxythiophene) for Chronic Neural Stimulation," *IEEE Trans. Neural Syst. Rehabil. Eng.*, vol. 15, pp. 502-508, 2007.
- [16] S. J. Wilks, S. M. Richardson-Burns, J. L. Hendricks, D. C. Martin, and K. J. Otto, "Poly(3,4-ethylenedioxythiophene) (PEDOT) as a micro-neural interface material for electrostimulation," *Front. Neuroengineering*, vol. 2, p. 7, 2009.
- [17] E. W. Keefer, B. R. Botterman, M. I. Romero, A. F. Rossi, and G. W. Gross, "Carbon nanotube coating improves neuronal recordings," *Nat. Nanotechnol.*, vol. 3, pp. 434-9, Jul 2008.
- [18] S. F. Cogan, A. A. Guzelian, W. F. Agnew, T. G. Yuen, and D. B. McCreery, "Over-pulsing degrades activated iridium oxide films used for intracortical neural stimulation," *J. Neurosci. Methods*, vol. 137, pp. 141-50, Aug 30 2004.
- [19] J. D. Weiland and D. J. Anderson, "Chronic neural stimulation with thin-film, iridium oxide electrodes," *IEEE Trans. Biomed. Eng.*, vol. 47, pp. 911-8, Jul 2000.
- [20] S. F. Cogan, "Neural Stimulation and Recording Electrodes," *Annu. Rev. Biomed. Eng.*, Apr 22 2008.